Antifungal Properties .- The fungistatic assay was carried out in Sabouraud's liquid (1 g of neopeptone Difco and 2 g of glucose Difco per 100 ml of distilled water) in 16×160 mm test tubes, each containing 4.5 ml of liquid medium which had been auto-claved at 120° for 15 min. The compounds to be tested were dissolved in 50% alcohol at initial concentration of 20 mg/ml. The solutions were subsequently diluted with sterile distilled water to give a concentration of 10 mg/ml. Successive decimal dilutions were made in distilled water. To tubes containing 4.5 ml of Sabouraud's liquid medium 0.5 ml of the solution of the drug was added, thereby obtaining concentrations of 1000, 500, 100, 10, and 1 μ g/ml of medium. Control tubes were prepared by adding 0.5 ml of distilled water to 4.5 ml of medium, alcohol being added to give concentrations identical with the tubes containing 1000 and 500 μ g of the drug. The filamentous fungi were incubated in Sabourand's agar at 25° for 2-3 weeks. A block of $2 \times 2 \times 2$ mm was then inoculated into the medium. All cultures were made in duplicate and were incubated at 25° for 14 days. Readings were then taken and were expressed as + if inhibition at the 100- μ g/ml level was complete and as ++ if total inhibition occurred at 10 µg/ml.

Results

The results are summarized in Table II. Clearly, compounds of type IIIa, b exhibit excellent fungistatic activity against several organisms. Optimum effect is achieved with lower esters 6 and 9. Lengthening of the chain results in diminished activity, notably against

TABLE II

ANTIFUNGAL ACTIVITIES

	N ∬∬
$R'O_2C'$	N_
	R

			$\underbrace{\text{Lowest level of}}_{T.}$		
			menta-		
			M.	gro-	T.
Compd	R	R′	can is	phytes	rubrum
2	l-Indanyl	CH3	+	+	+
4	l-Indanyl	C_2H_b	+	++	++
8	1-Tetralyl	CH3	+	+	++
6	1-Tetralyl	C2H5	++	++	++
9	1-Tetralyl	n-C3H7	++	++	++
10	l-Tetralyl	i-C3H7	+	+	+
11	l-Tetralyl	$n-C_4H_9$	+	++	++
12	1-Tetralyl	i-C4H9	+	++	++
13	1-Tetralyl	$n-C_{\delta}H_{ii}$	0	+	+
14	1-Tetralyl	$CH_{3}CHCH_{2}CH(CH_{3})_{2}$	0	0	0
15	1-Tetralyl	CH ₂ CH=CH ₂	+	++	++
16	l-Tetralyl	CH ₂ CH ₂ OCH ₃	+	++	++
17	1-Tetralyl	$CH_2CH_2OC_2H_5$	+	+	++
10	1	07	,		
18	I-Tetralyl		+	+	+
19	l-Tetralyl	C ₆ H ₁₁	0	0	++
20	l-Tetralyl	CH2C6H5	0	0	++
4 Tota	linhibition	of 100 um 1 + of 10		I	

Total inhibition at 100 μ g, +; at 10 μ g, ++.

Microsporum canis. Chain interruption, in one case, (13 vs. 17) increases activity. The introduction of bulky groups causes greatly diminished inhibition (*i.e.*, 18-20), while carboxylic acids 3 and 7 are totally inactive.

For comparative purposes sodium undecalenate, diamthazole,³ and chlormidazole⁴ were assayed concurrently against M. canis, Trichophyton mentagrophytes, and Trichophyton rubrum. None of these caused total inhibition below the 100-µg/ml level. One of our compounds, 1-(1-tetralyl)imidazole-5-carboxylic acid ethyl

(3) Asterol® (Roche).

(4) Polycid® (Grünenthal GinbH).

sented elsewhere.⁵

ester (6, proposed generic name, ethonamidate) has been selected for clinical evaluation. A more detailed pharmacological study of ethonanidate will be pre-

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Lincomycin. VII. 4'-Depropyl-4'-ethoxylincomycins

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Variation in the length of the 4'-alkyl substituent of the antibiotic lincomycin (1) produced a series of analogs, some of which possessed enhanced antibacterial activity.¹ This note describes the synthesis and antibacterial activity of 4'-depropyl-4'-ethoxylincomycin (3) and its *cis* isomer 4. These compounds may be classified as classical bioisosteres² of lincomycin and its cis isomer 2 in which the methylene adjacent to the proline ring in the 4'-propyl substituent is replaced by oxygen.

1-Carbobenzoxy-4-hydroxy-L-proline³ was converted to the benzyl ester (6) and the latter etherified by the excellent method of Kuhn,⁴ to form crude 7 in almost quantitative yield. Hydrogenolysis of 7 afforded 4ethoxy-L-proline (8) as well as a small amount of the diketopiperazine 15. Reductive methylation of 8 proceeded smoothly yielding 9. Condensation of 1methyl-4-ethoxy-L-proline (9) with methyl thiolincosaminide⁵ using the mixed-anhydride procedure led to crystalline 4'-depropyl-4'-ethoxylincomycin (3).

In a similar manner, 1-carbobenzoxy-4-cis-hydroxy-L-proline³ was converted to 1-methyl-4-cis-ethoxy-Lproline (14) and then to 4'-depropyl-4'-cis-ethoxylincomycin (4). In 4 the ethoxy substituent at C-4' is oriented *cis* to the L-amide group, the same configuration as *cis*-lincomycin.¹

4'-Depropyl-4'-ethoxylincomycin (3) and 4'-depropyl-4'-cis-ethoxylincomycin (4) have about 2%the activity of lincomycin when assayed in the standardcurve assay against Sarcina lutea.⁶ Both compounds were inactive when administered subcutaneously at

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320 mg/kg and orally at 800 mg/kg to mice infected with *Staphylococcus aureus*. Lincomycin, run as a control, protected at 13 mg/kg when administered subcutaneously and at 49 mg/kg orally.⁷

Experimental Section⁸

1-Carbobenzoxy-4-ethoxy-L-proline Benzyl Ester (7).— A mixture of 219 g of 1-carbobenzoxy-4-hydroxy-L-proline (5),³ 80 ml of benzyl alcohol, 300 ml of benzene, and 1 g of p-toluene-sulfonic acid was heated under reflux and the distillate was passed through a Dean–Stark distillation head. During 17 hr, an additional 400 mg of p-toluenesulfonic acid and 100 ml of benzene were added. The reaction mixture was separated into neutral and acidic fractions by partition between NaHCO₃ solution and benzene. The oily, neutral fraction weighed 219 g. Thin layer chromatography using a cyclohexane–acetone system (1:1) showed one spon at R_f 0.69 when sprayed with 0.1 N aqueous I₂ solution.

The crude ester from above was dissolved in 750 ml of dimethylformamide (DMF) and 372 g of EtI. Silver oxide (462 g) was added, and the mixture was stirred for 3 days at ambient temperature then filtered, and the residue was washed successively with DMF and CH₂Cl₂. The solvent was distilled under vacuum. The residue was dissolved in CH₂Cl₂. The solution was washed with H₂O several times and dried, and the solvent was distilled. The resulting oil, 223 g, showed one spot on the run as described above (R_t 0.76). 4-Ethoxy-L-proline (8) and 2,7-Diethoxyoctahydro-5H,10Hdipyrrolo[1,2-a:1,2'-d]pyrazine-5,10-dione (15).—Compound 7 (17 g) in 200 ml of MeOH containing 2 g of 10% Pd-C was shaken under H₂ for 18 hr. The catalyst was removed by filtration. The solvent was evaporated and the residue was crystallized from methanol-ether to yield 2.9 g of 8, mp 185-188°. The mother liquor was evaporated and the residue was dissolved in 150 ml of MeOH and rehydrogenated over 2 g of catalyst. An additional 2.2 g of 8, mp 194-198°, was thus obtained. An analytical sample was prepared by two recrystallizations from methanol-ether. It melted at 194-198° and showed one spot on the [methanol-NH₄OH (17%) 9.5(0.5); detection by minhydrid]; $[\alpha]n = 46\%$ (H₂O).

 $Anal. Caled for C_7H_{63}NO_3; C, 52.81; H, 8.23; N, 8.80. Found: C, 52.63; H, 8.46; N, 8.77.$

The nucleer liquors from some runs yielded a more soluble compound which was purified by recrystallization from ethyl acetate-Skellysolve B. Infrared analysis indicated the diketopiperazine **15**, mp 123-125°.

Anal. Caled for $C_{0}H_{22}N_{2}O_{3}$; C, 59.55; H, 7.85; N, 9.92, Found: C, 59.46; H, 7.92; N, 9.83.

1-Methyl-4-ethoxy-t-proline (9).—A mixture of 3.75 g of 8, 300 mg of 10% Pd-C, 3.5 ml of formalin, and 150 ml of MeOH was shaken under Π_2 for 4 hr. The catalyst was removed by fibration and the solveror was evaporated *in vacuo*. Berzene was added to the residue, nichydrin nest negative. The crude crystallive residue, nichydrin nest negative. The crude trystallive were dissolved in MeOH, and the solution was chrifted by fibration; crystals precipitated by the addition of ether. The yield of 9, np 109–113°, was 2.7 g. The analytical sample, np 117–119°, $(\alpha) \nu = 60^{\circ}$ (Π_2 O), was prepared by several recrystallizations from ethyl acetate.

Anal. Caled for $C_8H_{6.}NO_2$; C, 55.47; H, 8.73; N, 8.09, Found: C, 55.39; H, 8.85; N, 8.08.

4'-Depropyl-4'-ethoxylincomycin (3), -To a solution of 5.2 g of 1-methyl-4-ethoxy-1-proline (9) and 4.9 ml of $E_{1a}N$ in 200 ml of acercanirrile coded (act)° was added 4.1 ml of isoburyl chloroformate. The reaction mixture was stirred for 0.25 hr, and a solution of 10.95 g of methyl thiolineosaminide in 150 ml of water was added. The mixture was stirred at 0° for 1 hr and at ambient remperature for 7 hr. The acctanitrile was evaporated under vacuum. The residue was diluted with water and treated with two 30-g portions of Dowex 2 (OII form). The solution was lyophilized. The dried residue was shaken with 500 ml of acronoc and filtered. The acctone solution was acidified with dilute 11Cl, precipitating crude 3 HCl as a gum which crystallized on standing. It was recrystallized by dissolving in water and adding The yield of $3 \cdot \text{HCl}$ was 7.79 g (56.6%), mp 150–160° acctone. (solvated). Similar material on recrystallization afforded a hydrate, up $170 \cdot 175^{\circ}$ dec, $\{\alpha\}(e - 145)^{\circ}$ (H₂O).

4nal. Caled (or $C_6H_{32}N_2O_5$; C, 45.88; H, 7.48; N, 6.30; S, 7.21. Found: C, 45.84; H, 7.74; N, 6.05; S, 7.27 (corrected for 2.53% H₂O, Karl-Fischer ritration).

1-Carbobenzoxy-4-*cis*-ethoxy-1-proline Benzyl Ester (12). 1-Carbohenzoxy-4-allohydroxy-1-proline (10)³ (10 g) was esterified and ethylated as described for the preparation of **7**. The ernde product was chromatographed over 400 g of Florisil using Skellysolve B (10)-71°) with increasing amounts of acetone for elution. An eily fraction showing one spot by (1c)(R_{1} 0.74) was obtained and used directly in the following step.

4-*cis*-**Ethoxy**-**L**-**proline** (13).—In the manner described above for the preparation of 8, 3.4 g of 12 was hydrogenated to yield a total of 67.6% of 13, mp $185-205^\circ$, $[\alpha]n = -36^\circ$ (H₂t)). This material was difficult to crystallize, forming gels very readily. The melting point of the crystals obtained varied considerably, some melting as high as 214–216°.

Anal. Caled for C₇H₆₃NO₃: N, 8.80. Found: N, 8.44.

1-Methyl-4-c/s-ethoxy-L-proline (14),—In the manner described for the preparation of 9, 1 g of 13 was converted to 14 (900 mg), mp 163–165° (from McOII–EtOAc), { α |D = 61° (H₂O), Anal. Caled for C₈H₂NO₅: C, 55.46; H, 8.73; N, 8.09.

Found: C. 56.05; H, 9.02; N, 8.10. A partian was converted to the hydrochloride which was re-

crystallized from methanol other. It melted at 141-143°.

.1*nal.* Called for C₈11₆₅NO₈(11C1) C, 45.82; 11, 7.69; N, 6.69, Found: C, 46.00; 11, 7.52; N, 7.01.

4'-Depropyl-4'-*cis*-alloethoxylincomycin (4).—As previously described for the preparation of **3**, 0.52 g of **14** was treated with 1.09 g of methyl thioliacesaninide to yield 0.77 g (57.6%) of **4**,

⁽⁷⁾ C. Lewis, H. W. Clapp, and J. E. Gendy, rel 6, p 570.

⁽⁸⁾ Melting points were taken in a Thomas-Hoover Undateb apparatus and were corrected for stem exposure. This layer chromatography (ite) was carried out on adcrossibles coared with Brinkmann silica gel GF₃₃₄. Optical rotations were observed at 26° at $c \sim 1$. Infrared and num data are available out reades).

mp 146-152°. A portion was recrystallized from acetone-water.

It melted at 141–146° and had $[\alpha]_D + 132^\circ$ (H₂O). Anal. Calcd for C₁₇H₃₂N₂O₇S·HCl: C, 45.88; H, 7.48; N, 6.30; S, 7.21, Found: C, 45.39; H, 7.60; N, 6.41; S, 7.09 (corrected for 4.64% water, Karl-Fischer titration).

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Bis(pyrimidine nucleoside) Phosphates¹

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6-Mercaptopurine must be metabolized to its ribonucleotide before it can inhibit the growth of cells.² Cells that lack the IMP-GMP pyrophosphorylase necessary for this conversion are resistant to 6-mercaptopurine,² and to its ribonucleotide,³ which is unable to enter cells intact.⁴ In an effort to overcome this problem of resistance, a number of derivatives of 6-mercaptopurine ribonucleotide that might enter cells were synthesized.⁵ One of these, thioinosinyl- $(5' \rightarrow 5')$ thioinosine,⁵ was found to be cytotoxic to a subline of HEp-2 cells resistant to 6-mercaptopurine.³ This observation led us to synthesize the same type of derivative of the biologically active pyrimidine nucleoside analogs (1a-c and 6-azauridine). The same general approach used in our previous work⁵ was employed. 5-Bromo-2'-deoxyuridine (1b) and 2'-deoxy-5-iodouridine (1c) were converted to their 5'-O-trityl derivatives (2b and c) for acetylation. Removal of the trityl group from the acetylated nucleosides (3b and c) gave nucleosides (4b and c) suitably blocked for conversion to the desired phosphates. The preparation of 3'-Oacetyl-2'-deoxy-5-fluorouridine (4a) was described previously.⁵ 6-Azauridine was converted to its 2',3'-Oisopropylidene derivative (7).⁶ These blocked nucleosides (4 and 7) were then allowed to react with p-nitrophenylphosphorodichloridate⁷ and the blocked nucleotides (5) were treated with base to remove the acetyl group and the p-nitrophenyl group to give the desired 2'-deoxy-5-halouridylyl- $(5' \rightarrow 5')$ -2'-deoxy-5-halouridines (6).^{8,9} Treatment of the azauridine derivative (8) with base removed the *p*-nitrophenyl group, but removal of the 2',3'-O-isopropylidene group was easily accomplished by simply refluxing an aqueous solution of 9 which, in itself, is acidic enough to effect the hydrolysis to 10 (see Scheme I).

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Biologic Data.—The cytotoxic effect of these dinucleoside phosphates on KB cells is compared to the effect of the corresponding nucleosides in Table I. The cells were grown on glass and growth was measured by determination of protein content.¹⁰

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